

1 **Densely sampled viral trajectories suggest longer duration of acute infection with B.1.1.7**  
2 **variant relative to non-B.1.1.7 SARS-CoV-2**

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25  
26  
27 **Abstract.**

28  
29 To test whether acute infection with B.1.1.7 is associated with higher or more sustained nasopharyngeal viral concentrations, we assessed longitudinal PCR tests performed in a cohort of 65  
30 individuals infected with SARS-CoV-2 undergoing daily surveillance testing, including seven infected with B.1.1.7. For individuals infected with B.1.1.7, the mean duration of the proliferation  
31 phase was 5.3 days (90% credible interval [2.7, 7.8]), the mean duration of the clearance phase  
32 was 8.0 days [6.1, 9.9], and the mean overall duration of infection (proliferation plus clearance)  
33 was 13.3 days [10.1, 16.5]. These compare to a mean proliferation phase of 2.0 days [0.7, 3.3],  
34 a mean clearance phase of 6.2 days [5.1, 7.1], and a mean duration of infection of 8.2 days [6.5,  
35 9.7] for non-B.1.1.7 virus. The peak viral concentration for B.1.1.7 was 19.0 Ct [15.8, 22.0] compared to 20.2 Ct [19.0, 21.4] for non-B.1.1.7. This converts to 8.5 log<sub>10</sub> RNA copies/ml [7.6, 9.4]  
36 for B.1.1.7 and 8.2 log<sub>10</sub> RNA copies/ml [7.8, 8.5] for non-B.1.1.7. These data offer evidence that  
37 SARS-CoV-2 variant B.1.1.7 may cause longer infections with similar peak viral concentration  
38 compared to non-B.1.1.7 SARS-CoV-2. This extended duration may contribute to B.1.1.7 SARS-  
39 CoV-2's increased transmissibility.  
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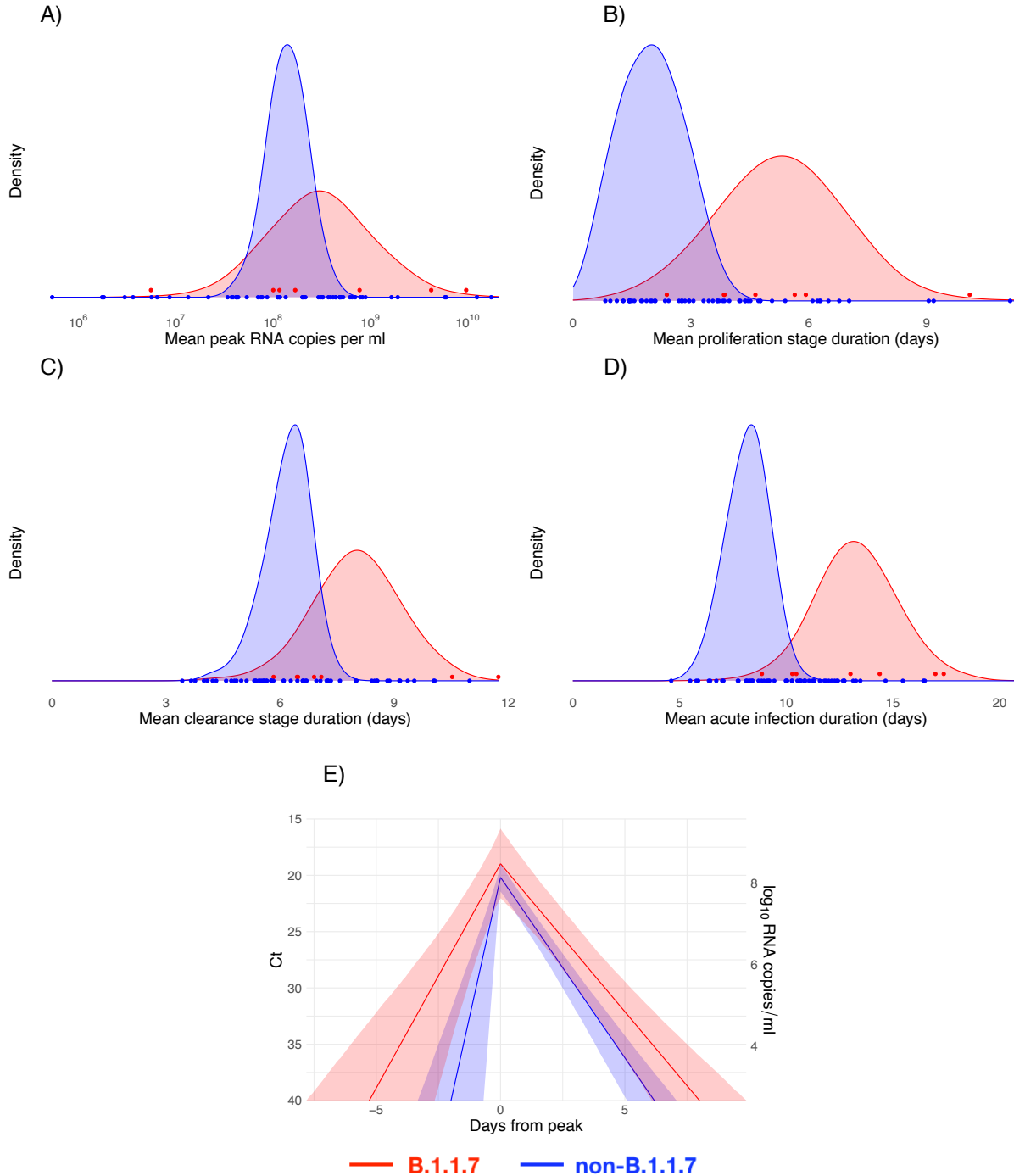
43 **Main text.**

44 The reasons for the enhanced transmissibility of SARS-CoV-2 variant B.1.1.7 are unclear. B.1.1.7  
45 features multiple mutations in the spike protein receptor binding domain<sup>1</sup> that may enhance ACE-  
46 2 binding<sup>2</sup>, thus increasing the efficiency of virus transmission. A higher or more persistent viral  
47 burden in the nasopharynx could also increase transmissibility. To test whether acute infection  
48 with B.1.1.7 is associated with higher or more sustained nasopharyngeal viral concentrations, we  
49 assessed longitudinal PCR tests performed in a cohort of 65 individuals infected with SARS-CoV-  
50 2 undergoing daily surveillance testing, including seven infected with B.1.1.7, as confirmed by  
51 whole genome sequencing.

52  
53 We estimated (1) the time from first detectable virus to peak viral concentration (proliferation time),  
54 (2) the time from peak viral concentration to initial return to the limit of detection (clearance time),  
55 and (3) the peak viral concentration for each individual (**Supplementary Appendix**).<sup>3</sup> We esti-  
56 mated the means of these quantities separately for individuals infected with B.1.1.7 and non-  
57 B.1.1.7 SARS-CoV-2 (**Figure 1**). For individuals infected with B.1.1.7, the mean duration of the  
58 proliferation phase was 5.3 days (90% credible interval [2.7, 7.8]), the mean duration of the clear-  
59 ance phase was 8.0 days [6.1, 9.9], and the mean overall duration of infection (proliferation plus  
60 clearance) was 13.3 days [10.1, 16.5]. These compare to a mean proliferation phase of 2.0 days  
61 [0.7, 3.3], a mean clearance phase of 6.2 days [5.1, 7.1], and a mean duration of infection of 8.2  
62 days [6.5, 9.7] for non-B.1.1.7 virus. The peak viral concentration for B.1.1.7 was 19.0 Ct [15.8,  
63 22.0] compared to 20.2 Ct [19.0, 21.4] for non-B.1.1.7. This converts to 8.5 log<sub>10</sub> RNA copies/ml  
64 [7.6, 9.4] for B.1.1.7 and 8.2 log<sub>10</sub> RNA copies/ml [7.8, 8.5] for non-B.1.1.7. Data and code are  
65 available online.<sup>4</sup>

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67 These data offer evidence that SARS-CoV-2 variant B.1.1.7 may cause longer infections with  
68 similar peak viral concentration compared to non-B.1.1.7 SARS-CoV-2, and this extended dura-  
69 tion may contribute to B.1.1.7 SARS-CoV-2's increased transmissibility. The findings are prelimi-  
70 nary, as they are based on seven B.1.1.7 cases. However, if borne out by additional data, a longer  
71 isolation period than the currently recommended 10 days after symptom onset<sup>5</sup> may be needed  
72 to effectively interrupt secondary infections by this variant. Collection of longitudinal PCR and test  
73 positivity data in larger and more diverse cohorts is needed to clarify the viral trajectory of variant  
74 B.1.1.7. Similar analyses should be performed for other SARS-CoV-2 variants such as B.1.351  
75 and P.1.

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**Figure 1. Estimated viral trajectories for B.1.1.7 and non-B.1.1.7 SARS-CoV-2.** Posterior distributions for the mean peak viral concentration (A), mean proliferation duration (B), mean clearance duration (C), mean total duration of acute infection (D), and mean posterior viral concentration trajectory (E) for the B.1.1.7 variant (red) and non-B.1.1.7 SARS-CoV-2 (blue). In (A)–(D), distributions depict kernel density estimates obtained from 2,000 draws from the posterior distributions for each statistic. Points depict the individual-level posterior means for each statistic. In (E), solid lines depict the estimated mean viral trajectory. Shaded bands depict the 90% credible intervals for the mean viral trajectory.

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137 **Supplementary Appendix.**

138  
139 Ethics.

140 Residual de-identified viral transport media from anterior nares and oropharyngeal swabs  
141 collected from players, staff, vendors, and associated household members from a professional  
142 sports league were obtained from BioReference Laboratories. In accordance with the guidelines  
143 of the Yale Human Investigations Committee, this work with de-identified samples was approved  
144 for research not involving human subjects by the Yale Internal Review Board (HIC protocol #  
145 2000028599). This project was designated exempt by the Harvard IRB (IRB20-1407).

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147 Study population. The data reported here represent a convenience sample including team staff,  
148 players, arena staff, and other vendors (e.g., transportation, facilities maintenance, and food  
149 preparation) affiliated with a professional sports league. Clinical samples were obtained by  
150 combined swabs of the anterior nares and oropharynx administered by a trained provider. Viral  
151 concentration was measured using the cycle threshold (Ct) according to the Roche cobas target  
152 1 assay. For an initial pool of 298 participants who first tested positive for SARS-CoV-2 infection  
153 during the study period (between November 28<sup>th</sup>, 2020 and January 20<sup>th</sup>, 2021), a diagnosis of  
154 “novel” or “persistent” infection was recorded. “Novel” denoted a likely new infection while  
155 “persistent” indicated the presence of virus in a clinically recovered individual. A total of 65  
156 individuals (90% male) had novel infections that met our inclusion criteria: at least five positive  
157 PCR tests (Ct < 40) and at least one test with Ct < 35. Seven of these individuals were infected  
158 with the B.1.1.7 variant as confirmed by genomic sequencing.

159  
160 Genome sequencing and lineage assignments: RNA was extracted from remnant  
161 nasopharyngeal diagnostic specimens and used as input for SARS-CoV-2 genomic sequencing  
162 as previously described.<sup>6</sup> Samples were sequenced on the Oxford Nanopore MinION. Consensus  
163 sequences were generated using the ARTIC Network analysis pipeline<sup>7</sup> and samples with >80%  
164 genome coverage were included in analysis. Individual SARS-CoV-2 genomes were assigned to  
165 PANGO lineages using Pangolin v.2.1.8.<sup>8</sup> All viral genomes assigned to the B.1.1.7 lineage were  
166 manually examined for representative mutations.<sup>9</sup>

167  
168 Converting Ct values to viral genome equivalents. To convert Ct values to viral genome  
169 equivalents, we first converted the Roche cobas target 1 Ct values to equivalent Ct values on a

170 multiplexed version of the RT-qPCR assay from the US Centers for Disease Control and  
171 Prevention.<sup>10</sup> We did this following our previously described methods.<sup>3</sup> Briefly, we adjusted the  
172 Ct values using the best-fit linear regression between previously collected Roche cobas target 1  
173 Ct values and CDC multiplex Ct values using the following regression equation:

$$y_i = \beta_0 + \beta_1 x_i + \epsilon_i$$

174  
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176  
177 Here,  $y_i$  denotes the  $i^{\text{th}}$  Ct value from the CDC multiplex assay,  $x_i$  denotes the  $i^{\text{th}}$  Ct value from the  
178 Roche cobas target 1 test, and  $\epsilon_i$  is an error term with mean 0 and constant variance across all  
179 samples. The coefficient values are  $\beta_0 = -6.25$  and  $\beta_1 = 1.34$ .

180  
181 Ct values were fitted to a standard curve in order to convert Ct value data to RNA copies. Synthetic  
182 T7 RNA transcripts corresponding to a 1,363 b.p. segment of the SARS-CoV-2 nucleocapsid gene  
183 were serially diluted from  $10^6$ - $10^0$  RNA copies/ $\mu\text{l}$  in duplicate to generate a standard curve<sup>11</sup>  
184 **(Supplementary Table 1)**. The average Ct value for each dilution was used to calculate the slope  
185 (-3.60971) and intercept (40.93733) of the linear regression of Ct on log-10 transformed standard  
186 RNA concentration, and Ct values from subsequent RT-qPCR runs were converted to RNA copies  
187 using the following equation:

$$\log_{10}([\text{RNA}]) = (Ct - 40.93733)/(-3.60971) + \log_{10}(250)$$

188  
189  
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191 Here, [RNA] represents the RNA copies /ml. The  $\log_{10}(250)$  term accounts for the extraction (300  
192  $\mu\text{l}$ ) and elution (75  $\mu\text{l}$ ) volumes associated with processing the clinical samples as well as the  
193 1,000  $\mu\text{l}/\text{ml}$  unit conversion.

#### 194 Model fitting.

195  
196 For the statistical analysis, we removed any sequences of 3 or more consecutive negative tests  
197 to avoid overfitting to these trivial values. Following our previously described methods,<sup>3</sup> we  
198 assumed that the viral concentration trajectories consisted of a proliferation phase, with  
199 exponential growth in viral RNA concentration, followed by a clearance phase characterized by  
200 exponential decay in viral RNA concentration.<sup>12</sup> Since Ct values are roughly proportional to the  
201 negative logarithm of viral concentration<sup>13</sup>, this corresponds to a linear decrease in Ct followed by  
202 a linear increase. We therefore constructed a piecewise-linear regression model to estimate the

203 peak Ct value, the time from infection onset to peak (*i.e.* the duration of the proliferation stage),  
 204 and the time from peak to infection resolution (*i.e.* the duration of the clearance stage). The  
 205 trajectory may be represented by the equation

$$E[Ct(t)] = \begin{cases} \text{l.o.d.} & t \leq t_o \\ \text{l.o.d.} - \frac{\delta}{t_p - t_o}(t - t_o) & t_o < t \leq t_p \\ \text{l.o.d.} - \delta + \frac{\delta}{t_r - t_p}(t - t_p) & t_p < t \leq t_r \\ \text{l.o.d.} & t > t_r \end{cases}$$

207  
 208  
 209 Here,  $E[Ct(t)]$  represents the expected value of the Ct at time  $t$ , “l.o.d” represents the RT-qPCR  
 210 limit of detection,  $\delta$  is the absolute difference in Ct between the limit of detection and the peak  
 211 (lowest) Ct, and  $t_o$ ,  $t_p$ , and  $t_r$  are the onset, peak, and recovery times, respectively.

212  
 213 Before fitting, we re-parametrized the model using the following definitions:

- 214
- 215 •  $\Delta Ct(t) = \text{l.o.d.} - Ct(t)$  is the difference between the limit of detection and the observed Ct
  - 216 value at time  $t$ .
  - 217 •  $\omega_p = t_p - t_o$  is the duration of the proliferation stage.
  - 218 •  $\omega_c = t_r - t_p$  is the duration of the clearance stage.

219  
 220 We constrained  $0.25 \leq \omega_p \leq 14$  days and  $2 \leq \omega_c \leq 30$  days to prevent inferring unrealistically small  
 221 or large values for these parameters for trajectories that were missing data prior to the peak and  
 222 after the peak, respectively. We also constrained  $0 \leq \delta \leq 40$  as Ct values can only take values  
 223 between 0 and the limit of detection (40).

224  
 225 We next assumed that the observed  $\Delta Ct(t)$  could be described the following mixture model:

$$226 \Delta Ct(t) \sim \lambda \text{Normal}(E[\Delta Ct(t)], \sigma(t)) + (1 - \lambda) \text{Exponential}(\log(10)) \Big|_0^{\text{l.o.d}}$$

227  
 228  
 229 where  $E[\Delta Ct(t)] = \text{l.o.d.} - E[Ct(t)]$  and  $\lambda$  is the sensitivity of the q-PCR test, which we fixed at 0.99.  
 230 The bracket term on the right-hand side of the equation denotes that the distribution was truncated  
 231 to ensure Ct values between 0 and the limit of detection. This model captures the scenario where

232 most observed Ct values are normally distributed around the expected trajectory with standard  
233 deviation  $\sigma(t)$ , yet there is a small (1%) probability of an exponentially distributed false negative  
234 near the limit of detection. The  $\log(10)$  rate of the exponential distribution was chosen so that 90%  
235 of the mass of the distribution sat below 1 Ct unit and 99% of the distribution sat below 2 Ct units,  
236 ensuring that the distribution captures values distributed at or near the limit of detection. We did  
237 not estimate values for  $\lambda$  or the exponential rate because they were not of interest in this study;  
238 we simply needed to include them to account for some small probability mass that persisted near  
239 the limit of detection to allow for the possibility of false negatives.

240  
241 We used a hierarchical structure to describe the distributions of  $\omega_p$ ,  $\omega_r$ , and  $\delta$  for each individual  
242 based on their respective population means  $\mu_{\omega_p}$ ,  $\mu_{\omega_r}$ , and  $\mu_{\delta}$  and population standard deviations  
243  $\sigma_{\omega_p}$ ,  $\sigma_{\omega_r}$ , and  $\sigma_{\delta}$  such that

244  
245  $\omega_p \sim \text{Normal}(\mu_{\omega_p}, \sigma_{\omega_p})$

246  $\omega_r \sim \text{Normal}(\mu_{\omega_r}, \sigma_{\omega_r})$

247  $\delta \sim \text{Normal}(\mu_{\delta}, \sigma_{\delta})$

248  
249 We inferred separate population means ( $\mu_{\cdot}$ ) for B.1.1.7- and non-B.1.1.7-infected individuals. We  
250 used a Hamiltonian Monte Carlo fitting procedure implemented in Stan (version 2.24)<sup>14</sup> and R  
251 (version 3.6.2)<sup>15</sup> to estimate the individual-level parameters  $\omega_p$ ,  $\omega_r$ ,  $\delta$ , and  $t_p$  as well as the  
252 population-level parameters  $\sigma^*$ ,  $\mu_{\omega_p}$ ,  $\mu_{\omega_r}$ ,  $\mu_{\delta}$ ,  $\sigma_{\omega_p}$ ,  $\sigma_{\omega_r}$ , and  $\sigma_{\delta}$ . We used the following priors:

253  
254 *Hyperparameters:*

255  
256  $\sigma^* \sim \text{Cauchy}(0, 5) [0, \infty]$

257  
258  $\mu_{\omega_p} \sim \text{Normal}(14/2, 14/6) [0.25, 14]$

259  $\mu_{\omega_r} \sim \text{Normal}(30/2, 30/6) [2, 30]$

260  $\mu_{\delta} \sim \text{Normal}(40/2, 40/6) [0, 40]$

261  
262  $\sigma_{\omega_p} \sim \text{Cauchy}(0, 14/\tan(\pi(0.95-0.5))) [0, \infty]$

263  $\sigma_{\omega_r} \sim \text{Cauchy}(0, 30/\tan(\pi(0.95-0.5))) [0, \infty]$

264  $\sigma_{\delta} \sim \text{Cauchy}(0, 40/\tan(\pi(0.95-0.5))) [0, \infty]$



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*Individual-level parameters:*

$$\omega_p \sim \text{Normal}(\mu_{\omega_p}, \sigma_{\omega_p}) [0.25, 14]$$

$$\omega_r \sim \text{Normal}(\mu_{\omega_r}, \sigma_{\omega_r}) [2, 30]$$

$$\delta \sim \text{Normal}(\mu_{\delta}, \sigma_{\delta}) [0, 40]$$

$$t_p \sim \text{Normal}(0, 2)$$

The values in square brackets denote truncation bounds for the distributions. We chose a vague half-Cauchy prior with scale 5 for the observation variance  $\sigma^*$ . The priors for the population mean values ( $\mu$ ) are normally distributed priors spanning the range of allowable values for that parameter; this prior is vague but expresses a mild preference for values near the center of the allowable range. The priors for the population standard deviations ( $\sigma$ ) are half Cauchy-distributed with scale chosen so that 90% of the distribution sits below the maximum value for that parameter; this prior is vague but expresses a mild preference for standard deviations close to 0.

We ran four MCMC chains for 1,000 iterations each with a target average proposal acceptance probability of 0.8. The first half of each chain was discarded as the warm-up. The Gelman R-hat statistic was less than 1.1 for all parameters. This indicates good overall mixing of the chains. There were no divergent iterations, indicating good exploration of the parameter space. The posterior distributions for  $\mu_{\delta}$ ,  $\mu_{\omega_p}$ , and  $\mu_{\omega_r}$ , were estimated separately for individuals infected with B.1.1.7 and non-B.1.1.7. These are depicted in **Figure 1** (main text). Draws from the individual posterior viral trajectory distributions are depicted in **Supplementary Figure 1**. The mean posterior viral trajectories for each individual are depicted in **Supplementary Figure 2**.

Checking for influential outliers. To examine whether the posterior distributions for the B.1.1.7-infected individuals reflected the influence of a single outlier, we re-fit the model seven times, omitting one of the B.1.1.7 trajectories each time. The inferred parameter values were fairly consistent, though omitting either of two of the B.1.1.7 cases (cases 5 and 6 in **Supplementary Table 2**). yields an infection duration with a 90% credible interval that overlaps with that of the non-B.1.1.7 90% credible interval for infection duration.

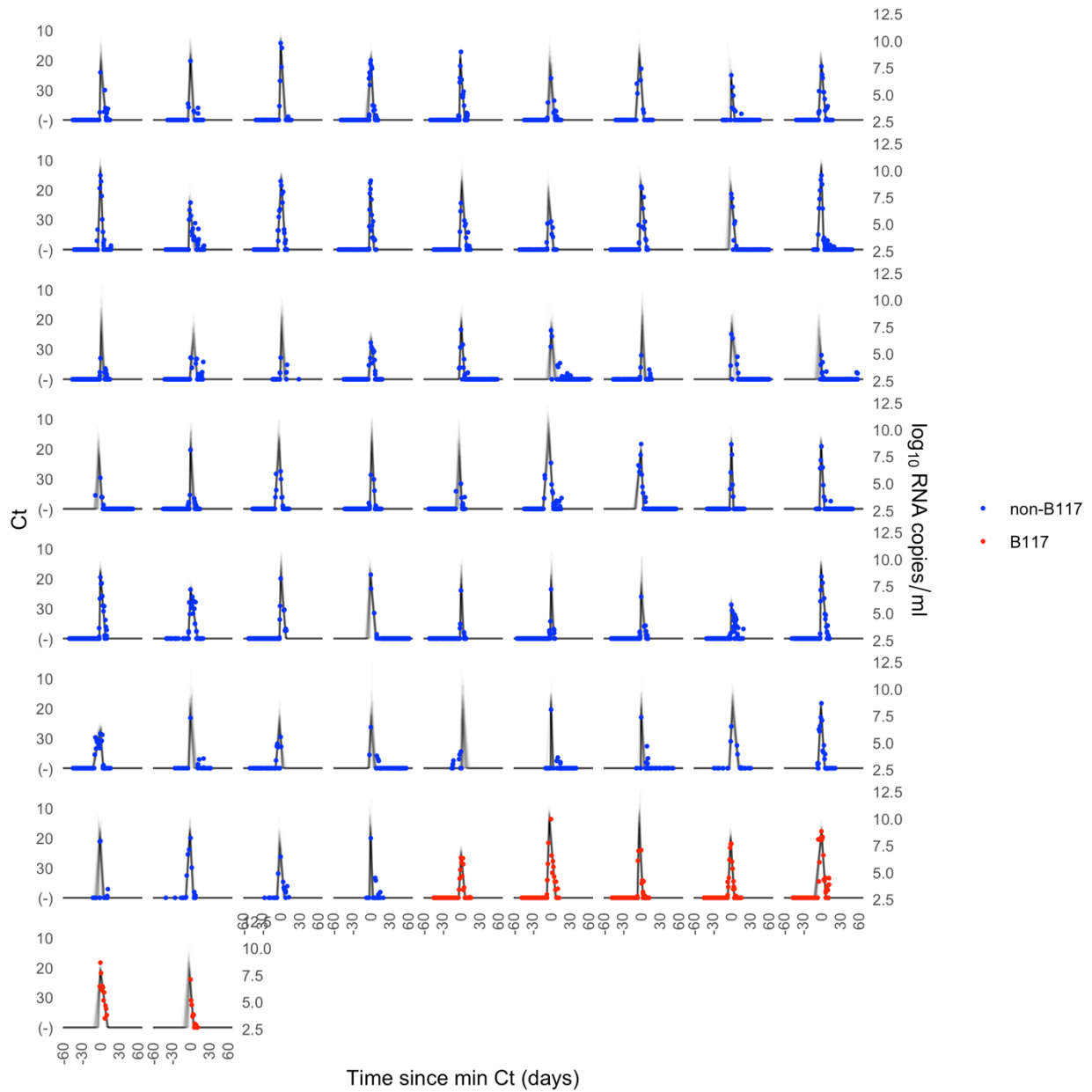
| Standard<br>(copies/ul) | Replicate 1 (Ct) | Replicate 2 (Ct) | Average Ct |
|-------------------------|------------------|------------------|------------|
| 10 <sup>6</sup>         | 19.3             | 19.7             | 19.5       |
| 10 <sup>5</sup>         | 23.0             | 21.2             | 22.1       |
| 10 <sup>4</sup>         | 26.9             | 26.7             | 26.8       |
| 10 <sup>3</sup>         | 30.6             | 30.4             | 30.5       |
| 10 <sup>2</sup>         | 34.0             | 34.0             | 34.0       |
| 10 <sup>1</sup>         | 37.2             | 36.6             | 36.9       |
| 10 <sup>0</sup>         | N/A              | 39.9             | 39.9       |

298 **Supplementary Table 1. Standard curve relationship between virus RNA copies and Ct values.** Synthetic T7  
 299 RNA transcripts corresponding to a 1,363 base pair segment of the SARS-CoV-2 nucleocapsid gene were serially  
 300 diluted from 10<sup>6</sup>-10<sup>0</sup> and evaluated in duplicate with RT-qPCR. The best-fit linear regression of the average Ct on the  
 301 log<sub>10</sub>-transformed standard values had slope -3.60971 and intercept 40.93733 (R<sup>2</sup> = 0.99).

| Omitted B117 Case            | Proliferation duration (days) [90% CI] | Clearance duration (days) [90% CI] | Infection duration (days) [90% CI] | Peak viral concentration (log(copies/ml)) [90% CI] |
|------------------------------|--|------------------------------------|------------------------------------|--|
| None                         | 5.3 [2.7, 7.8]                         | 8.0 [6.1, 9.9]                     | 13.3 [10.1, 16.5]                  | 8.5 [7.6, 9.4]                                     |
| 1                            | 5.5 [3.0, 8.1]                         | 8.3 [6.3, 10.3]                    | 13.9 [10.6, 17.0]                  | 8.8 [7.9, 9.8]                                     |
| 2                            | 5.7 [3.1, 8.4]                         | 7.5 [5.1, 9.6]                     | 13.2 [9.8, 16.5]                   | 8.2 [7.4, 9.1]                                     |
| 3                            | 5.9 [3.3, 8.6]                         | 8.3 [6.3, 10.3]                    | 14.2 [11.0, 17.4]                  | 8.2 [7.4, 9.1]                                     |
| 4                            | 5.4 [2.7, 7.9]                         | 8.5 [6.3, 10.5]                    | 13.9 [10.5, 17.0]                  | 8.5 [7.6, 9.4]                                     |
| 5                            | 4.3 [1.8, 6.9]                         | 8.3 [6.2, 10.3]                    | 12.6 [9.4, 15.8]                   | 8.4 [7.5, 9.3]                                     |
| 6                            | 5.4 [3.0, 7.9]                         | 7.1 [5.1, 9.1]                     | 12.6 [9.4, 15.6]                   | 8.6 [7.8, 9.6]                                     |
| 7                            | 5.2 [2.6, 7.7]                         | 8.1 [6.0, 10.2]                    | 13.3 [10.1, 16.6]                  | 8.6 [7.8, 9.4]                                     |
| <b>Non-B.1.1.7 reference</b> | 2.0 [0.7, 3.3]                         | 6.2 [5.1, 7.1]                     | 8.2 [6.5, 9.7]                     | 8.2 [7.8, 8.5]                                     |

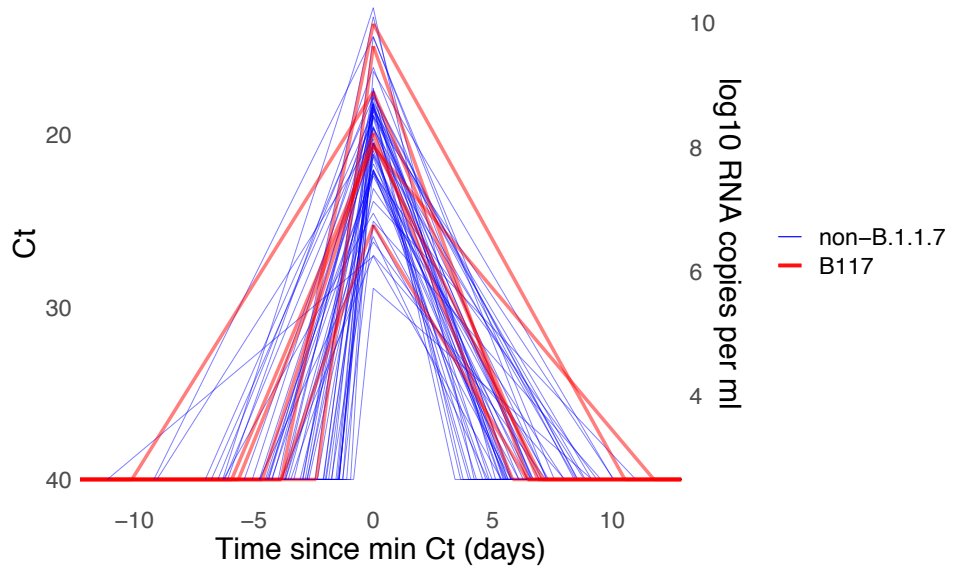
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**Supplementary Table 2. Posterior population mean viral trajectory parameter values and 90% credible intervals for B.1.1.7 infections when omitting single trajectories.** Each row corresponds to a model fit obtained by omitting one person who was infected with B.1.1.7, so that the parameter values are informed by six of the seven B.1.1.7 infections. The final row lists the fitted parameter values for the non-B.1.1.7 infections for reference.



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**Supplementary Figure 1. Ct values for 65 individuals with estimated viral trajectories.** Each pane depicts the recorded Ct values (points) and derived log-10 genome equivalents per ml ( $\log_{10}(ge/ml)$ ) for a single person during the study period. Points along the horizontal axis represent negative tests. Time is indexed in days since the minimum recorded Ct value (maximum viral concentration). Individuals with confirmed B.1.1.7 infections are depicted in red. Non-B.1.1.7 infections are depicted in blue. Lines depict 100 draws from the posterior distribution for each person's viral trajectory.



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**Supplementary Figure 2. Mean posterior viral trajectories for each person in the study.** Lines depict the posterior mean viral trajectory specified by the posterior mean proliferation time, mean clearance time, and mean peak Ct. Trajectories are aligned temporally to have the same peak time. B.1.1.7 trajectories are depicted in red, non-B.1.1.7 in blue.